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REFERENCES

- [1] Pulte D, Castro FA, Jansen L, et al. Trends in survival of chronic lymphocytic leukemia patients in Germany and the USA in the first decade of the twenty-first century. *J Hematol Oncol*. 2016;9:28

- [2] Falchi L, Keating MJ, Wang X, et al. Clinical characteristics, response to therapy, and survival of African American patients diagnosed with chronic lymphocytic leukemia: Joint experience of the MD Anderson cancer center and Duke university medical center. *Cancer*. 2013;119:3177–3185.
- [3] Vasylyev A, Loginov A, Molostvova V, et al. Prevalence and cumulative 5-year incidence of chronic lymphocytic leukemia in the adult population in the Russian Federation and Ukraine: Data from the leu-kospect study. *Hematology*. 2017;22:16–24.
- [4] Wu SJ, Chiang CJ, Lin CT, et al. Improving but inferior survival in patients with chronic lymphocytic leukemia in Taiwan: A population-based study, 1990–2004. *PLoS One*. 2013;8:e62930.
- [5] Hallek M, Cheson BD, Catovsky D, et al. Guidelines for the diagnosis and treatment of chronic lymphocytic leukemia: A report from the international workshop on chronic lymphocytic leukemia updating the national cancer institute-working group 1996 guidelines. *Blood*. 2008;111:5446–5456.
- [6] World Health Statistics 2016: Monitoring health for the SDGs. http://www.who.int/gho/publications/world_health_statistics/2016/en/ Accessed March 17, 2017.
- [7] Mato A, Nabhan C, Kay NE, et al. Real-world clinical experience in the connect(R) chronic lymphocytic leukaemia registry: A prospective cohort study of 1494 patients across 199 US centres. *Br J Haematol*. 2016;175:892–903.
- [8] Rodrigues CA, Goncalves MV, Ikoma MR, et al. Diagnosis and treatment of chronic lymphocytic leukemia: Recommendations from the Brazilian group of chronic lymphocytic leukemia. *Rev Bras Hematol Hemoter*. 2016;38:346–357.

SUPPORTING INFORMATION

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Urinary prednisolone excretion is a determinant of serum hepcidin levels in renal transplant recipients

To the Editor:

Hepcidin, which is synthesized and secreted by the liver, is considered the master regulator of iron homeostasis. Hepcidin regulates the amount of iron absorbed from the intestines and the iron release from the reticuloendothelial system by degrading ferroportin, the iron transporter located at the duodenal enterocytes and macrophages. Circulating levels of hepcidin are known to be controlled by available iron stores, inflammation, hypoxia, insulin levels, and erythropoiesis.

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TABLE 1 Baseline characteristics of 551 renal transplant recipients according to tertiles of 24-h urinary prednisolone excretion

Variables	All patients	Tertiles of urinary prednisolone excretion (pmol/24 h)			P value
		T1	T2	T3	
Urinary prednisolone (pmol/24 h)	758 (371–1278)	256 (125–371)	755 (619–904)	1595 (1273–2325)	<.001
General characteristics					
Age (years)	51 ± 12	53 ± 12	53 ± 12	49 ± 12	.003
Male sex (n %)	298 (54)	109 (60)	107 (58)	82 (45)	.006
eGFR (mL/min/1.73 m ²)	47 ± 16	41 ± 16	46 ± 15	54 ± 13	<.001
Laboratory parameters					
Hepcidin (ng/mL)	7.2 (3.3–13.5)	8.6 (4.3–14.3)	7.0 (2.8–14.3)	5.8 (2.8–11.4)	.003
Hemoglobin (g/dL)	13.8 ± 1.6	13.6 ± 1.6	13.8 ± 1.6	14.2 ± 1.4	<.001
Ferritin (g/L)	156.0 (80.0–283.0)	189 (102–316)	163 (77–288)	122 (68–233)	.001
EPO (IU/L)	17.4 (12.0–24.3)	17.7 (11.8–25.4)	17.9 (13.0–24.3)	16.6 (11.4–22.9)	.28
hs-CRP (mg/L)	2.0 (0.8–4.8)	2.1 (1.0–5.1)	2.7 (0.9–4.8)	1.7 (0.7–3.6)	.06
Insulin (μU/mL)	11.1 (7.9–16.3)	11.4 (7.8–17.5)	11.2 (8.3–15.1)	10.8 (7.8–15.8)	.53

eGFR, estimated glomerular filtration rate; EPO, erythropoietin; hs-CRP, high sensitivity C-reactive protein.

Recently, hepcidin antagonists have been introduced as potential treatment to improve iron-restrictive anemia. By improving iron availability and subsequently hemoglobin levels, hepcidin antagonists might be able to improve quality of life and outcome in different patient settings. Therefore, all factors that affect serum hepcidin levels are clinically relevant specifically in populations where in the future the use of hepcidin antagonists may be considered. It has already been established that both testosterone¹ and estrogens² are associated with suppression of serum hepcidin in men. On the other hand progesterone, the anabolic steroid epitostanol, as well as the progesterone antagonist, mifepristone, are able to induce hepcidin biosynthesis in a zebrafish model.³

Synthetic glucocorticoids, like prednisone, and its active metabolite prednisolone are often used in immunosuppressive regimens for renal transplant recipients (RTRs). To date, possible effects of these synthetic glucocorticoids on serum hepcidin levels in humans are unknown. Here, we report on the association of serum hepcidin with 24-h urinary prednisolone excretion, as a measure of 24-h prednisolone exposure.

For this study, 606 stable RTRs with a functioning graft beyond the first year after transplantation were included.⁴ All RTRs gave written informed consent for the study and approval by institutional review board was obtained (METc 2001/039). For the current analyses, we excluded patients with missing data on serum hepcidin ($n = 45$) and urinary prednisolone excretion ($n = 10$), resulting in 551 RTRs eligible for analyses. Serum hepcidin-25 was assessed by dual-monoclonal sandwich ELISA immunoassay. Urinary prednisolone measurements were carried out with validated high-performance liquid chromatography assay with diode-array detection after extraction with diethylether. Renal function was assessed by estimating glomerular filtration rate (eGFR) applying the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) equation.

IBM SPSS Statistics 23 was used for statistical analyses. For baseline characteristics, one-way ANOVA, Kruskal-Wallis test, and Chi square test were applied, as appropriate. Furthermore, univariate linear regression analysis was performed to assess the determinants of serum hepcidin, followed by multivariable linear regression analysis with stepwise

backward procedure. P values for inclusion and exclusion were set at 0.20 and 0.10, respectively. Variables with a skewed distribution were natural log transformed for analyses. Separate linear regression analysis was performed to assess the association between serum hepcidin and hemoglobin level in RTRs after adjustment for age, sex, and eGFR.

Mean age of the 551 RTRs was 51 ± 12 years with 55% being men. Patients were included at median 6.0 (interquartile range [IQR], 2.6–11.6) years after transplantation. All RTRs used prednisolone orally ranging from 5 to 10 mg once daily. Median 24-h urinary prednisolone excretion was 758 (371–1278) pmol/24 h. The association of the individual daily prednisolone dose with 24-h urinary prednisolone excretion was $\beta = 0.23$, $P < .001$. Across tertiles of 24-h urinary prednisolone excretion, RTRs within the highest tertile of prednisolone excretion were younger, less frequently men, had lower systolic blood pressure and higher estimated glomerular filtration rate (eGFR) compared to those within the other two tertiles. Furthermore, higher hemoglobin, lower hepcidin, and lower ferritin levels were noted in RTRs in the highest tertile of prednisolone excretion compared to RTRs in the other two tertiles (Table 1).

In univariate regression analysis, serum hepcidin was found to be negatively associated with 24-h urinary prednisolone excretion ($\beta = -0.15$, $P < .001$). After adjustment for high-sensitivity C-reactive protein (hs-CRP), hepcidin remained associated with prednisolone ($\beta = -0.13$, $P = .002$). Further adjustment for eGFR did not materially alter this association ($\beta = -0.12$, $P = .006$). When including prednisolone in a backward multivariate model with age, sex, eGFR, hs-CRP, ferritin, hemoglobin, erythropoietin, and insulin, prednisolone remained an independent determinant of hepcidin ($\beta = -0.10$, $P = .001$), besides expected relationships of hepcidin with ferritin, hs-CRP, erythropoietin and insulin. In addition, serum hepcidin was found to be a determinant of hemoglobin levels ($\beta = -0.08$, $P = .03$) independently of age, sex, and eGFR.

In this study, we show that 24-h urinary prednisolone excretion is negatively associated with serum hepcidin in RTRs irrespective of potential confounders, including eGFR. All RTRs in our cohort used a low dose

prednisolone (5–10 mg/day). Remarkably, this resulted in a broad range of 24-h urinary prednisolone excretion and a modest association with the daily prednisolone dose, in keeping with considerable inter-subject pharmacokinetic variability.⁵ Twenty-4 h urinary prednisolone excretion is considered to reflect the overall exposure to prednisolone. Previously, it has been shown that prednisolone dose-dependently inhibits the release of interleukin-6 (IL-6)⁶ which is known to induce hepcidin expression. We had no data available on IL-6 levels to assess whether effects on IL-6 is the mechanism behind the association of prednisolone with hepcidin. The possible role of prednisolone as a direct hepcidin antagonist and possible mechanisms linking prednisolone with hepcidin need to be delineated in more detail in future studies.

The major strength of this report is the large cohort of RTRs with availability of concurrent 24-h urinary prednisolone excretion and hepcidin data. Limitations are that it comprises a single center study, and that we cannot exclude the possibility of residual confounding.


In conclusion, lower serum hepcidin levels are related to higher 24-h urinary prednisolone excretion in RTRs independent of clinically relevant covariates. Our findings extend earlier data concerning effects of other (synthetic) steroids on hepcidin regulation, and provide a rationale to more precisely delineate direct or indirect effects of glucocorticoids on hepcidin regulation. From a clinical perspective, our findings lend support to the possibility that prednisolone may be regarded as a hitherto unappreciated hepcidin antagonist.

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CONFLICT OF INTEREST

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REFERENCES

- [1] Bachman E, Feng R, Travison T, et al. Testosterone suppresses hepcidin in men: a potential mechanism for testosterone-induced erythrocytosis. *J Clin Endocrinol Metab.* 2010;95:4743–4747.
- [2] Lehtihet M, Bonde Y, Beckman L, et al. Circulating hepcidin-25 is reduced by endogenous estrogen in humans. *PLoS One.* 2016;11:e0148802.
- [3] Li X, Rhee DK, Malhotra R, et al. Progesterone receptor membrane component-1 regulates hepcidin biosynthesis. *J Clin Invest.* 2016;126:389–401.
- [4] Zelle DM, Corpeleijn E, van Ree RM, et al. Markers of the hepatic component of the metabolic syndrome as predictors of mortality in renal transplant recipients. *Am J Transplant.* 2010;10:106–114.
- [5] Ionita IA, Ogasawara K, Gohh RY, Akhlaghi F. Pharmacokinetics of total and unbound prednisone and prednisolone in stable kidney transplant recipients with diabetes mellitus. *Ther Drug Monit.* 2014;36:448–455.
- [6] de Kruif MD, Lemaire LC, Giebelen IA, et al. Prednisolone dose-dependently influences inflammation and coagulation during human endotoxemia. *J Immunol.* 2007;178:1845–1851.

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Characterization of *TP53* mutations in clonal cytopenia of undetermined significance

To the Editor:

The diagnosis of myelodysplastic syndrome (MDS) requires persistent cytopenia with at least one of the following criteria: dysplasia in at least 10% of cells in any hematopoietic lineage, increased myeloblasts (5–19%) in bone marrow (or 2–19% myeloblast in peripheral blood), or MDS defining cytogenetic abnormalities. Some patients have cytopenia and/or gene mutations, but do not meet other criteria of MDS.¹ These pre-MDS conditions include idiopathic cytopenia of undetermined significance (ICUS), clonal hematopoiesis of indeterminate potential (CHIP) and clonal cytopenia of undetermined significance (CCUS). The mutations frequently identified in these pre-MDS conditions, including *DNMT3A*, *TET2*, and *ASXL1*, are also the common mutations detected in MDS.² ICUS, CHIP, and CCUS all carry an increased risk for progression to MDS. The rate of progression to MDS varies, likely depending on the specific genes that are mutated and their mutation burden. The role of each individual mutation in disease progression is not well characterized.

TP53 is a tumor suppressor gene that has been studied extensively in MDS and AML, in which the mutations are associated with a complex karyotype and a poor prognosis. Its mutations also occur in CHIP and CCUS.^{2,3} The characteristics of *TP53* mutations and their role in disease progression in these pre-MDS conditions are unknown. In this study, we aim to characterize the clinicopathological features of CCUS cases associated with *TP53* mutations.